Histological Interpretation of \textit{Helicobacter pylori} Density Relevant to Gastric Pathology and its Bias

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Abstract

A lot of articles related to \textit{Helicobacter pylori} (HP) have been produced from bench to bedside about active gastritis, gastric ulcer, duodenal ulcer and even gastric malignancies. Thanks to the efforts of eradication of HP, the necessity of accepting surgical intervention for aggravated peptic ulcer has been reduced obviously. The battle between human being and HP keeps going on from eradication of HP infection to optimal antibiotic administration to prevent progressive drug resistance. Now we will bring out our differentiated pathologic findings and long term clinical experience to deal with the limitations of histological examination and correlate with real clinical situation, moreover make evidence based therapeutic choice.

Keywords: Gastro duodenal mucosa; Giemsa stain; \textit{Helicobacter pylori}; Immunohistochemical stain

Introduction

\textit{Campylobacter pylori} (\textit{Helicobacter pylori}) has been identified as a gastric pathogen by Barry J Marshall and J Robin Warren since 1982 [1]. \textit{Helicobacter pylori} (HP) indwells in the mucous film on gastric mucosal surface and superficial gastric pits to escape from high acidity in stomach [1-4]. In our observation, HP infection can be found from gastric cardia to pylorus but very rarely found in duodenum even in the gastric metaplasia site or duodenum, though Barrett’s esophagus and duodenal bulb to descending duodenum could find HP mentioned in the Maastricht V/Florence Consensus Report [4]. Costs, sensitivity and specificity of various detection methods, check up facility and techniques for HP detection are well documented [5-9]. A histopathologic exam for HP density needs precise interpretation, or it will affect the sensitivity and specificity of HP detection [5,10,11]. Here in we would like to offer some findings and discussion for sharing experience.

Material and Methods

The HP density in gastric pits and on mucosal surface are detected through H & E stain (Figures 1A & 1B), Giemsa stain (Figures 2A - 2C) (Giemsa’s stain solution of Muto Pure Chemicals Co., Ltd) or Immunohistochemical (IHC) stain (primary antibody: Zytomed Rabbit anti-\textit{Helicobacter pylori} polyclonal antibody, and Bond Polymer Re- fine Detection kit of Leica Co.) under microscopy. H & E stain necessitates 400X high power view to confirm HP density. Although Giemsa stain can enhance the contrast of HP staining, HP infection must be checked at 200X to 400X high power field to distinguish the shape of curved rod or coccoid form, unlike the IHC staining that brown signals of HP density can be read at 100X to 200X magnifications without difficulty.
The gastric tissue can also be put in *Campylobacter* Like Organism (CLO) test kit to detect the presence of HP. The HP densities can be categorized into high, medium, low and trace levels (Figures 3A-3C).

**Results and Discussion**

Presence of HP density is frequently associated with erosive gastritis, gastric ulcer, active gastritis [11,12], but not vice versa, nor absolute proportional relationship (Figure 3C). The inflammatory situation may be affected by individual genetic factor at least [13]. In our observation, atrophic gastritis and intestinal metaplasia do not exclusively coexist with HP infection in submitted mucosal tissue specimen. Incidental use of antibiotics within 3 weeks before histologic exam of HP infection may lower the positive rate. HP infection rate seems decreased among some hospitals in Taiwan like some other areas in the world [5,14].

The fundic glands polyp have markedly lower HP densities (Figure 2C) comparing with those of the remaining gastric biopsy specimens in the same hospital [15], like the rarity of intestinal metaplasia in fundic gland polyp. Intestinal metaplasia (Figure 4) and fundic gland polyp will push aside the HP density, and the biopsies on these lesions without HP density should not exclude the presence of...
HP over the remaining gastric mucosal areas. Severe ulcerative gastric lesion at least will lose original HP density on account of partial erosion to total loss of mucosal layer, and may harbor nonspecific bacterial (Figures 3A & 5B) or even fungal colonization due to loss of local immunity.

‘Trace’ HP density can be a problem of pathological diagnosis, because it could be an easy way for interpreting unconfirmed HP morphology or a time consuming job to confirm a trace HP density but get an uncertain clinical significance for HP eradication treatment. Anyway, precipitates or striae of mucous covering without regular bacterial contour, and neither gray staining for H & E stain nor dark blue staining for Giemsa stain should not be interpreted as low or trace HP density. Otherwise, these over interpretations of so called gold standard might reflect in appropriate lower sensitivity of other HP tests such as stool antigen test in some nations [16,17] comparing with that of another nation [18]. More expensive HP Immunohistochemical (IHC) stain can reduce false positive or false negative results dramatically [19]. Bacterial contamination in duodenal biopsy may also occur from the HP in gastric secretion or nonspecific bacteria on oropharyngeal mucosa. These false trace HP finding could be proved with 13C-urea Breath test [5], which detects more carbon dioxide breathed out after ingestion of urea-containing tablet. Medium HP density or high HP density has less controversy on pathological interpretation.

Endoscopic check up for upper gastrointestinal tract is routine for most hospitals. Tissue examination with CLO test is cheap and convenient, although for some reasons there are discrepant results comparing with the H & E stain or Giemsa’s stain [20], that are partly caused by recognizing contaminated oropharyngeal bacterial as HP (Figures 6A-6E) or neglecting low HP prevalence by interpreter [21,22]. Nevertheless, false negative of invasive procedure for tissue examination related to sampling error from patchy HP distribution does occur (Figure 3A2) [5,23]. Additional test for any single negative or incompatible ‘tissue’ finding can improve the detection quality and interpretation experience via developed noninvasive tests for HP infection.
Figure 6B (1,2): Giemsa stain shows contaminated cocci (arrows) in B1 and contaminated bacilli (arrows) in B2. We can see nearby oropharyngeal squamous epithelial cell (arrow head).

Figure 6C (1,2): HP immunostain on the large coccal bacterial clumps (arrows) show only partially or weakly positive immunostaining, and proved to be oropharyngeal contamination (400X).

Figure 6D: Some contaminated bacteria can be pressed onto the mucosal surface intimately (short arrow) by biopsy forceps. However, the other part of contaminated bacteria and red foreign bodies are away from the mucosal surface (long arrow).

Figure 6E: Sharp edges on the tip of endoscope may scrape off some oropharyngeal bacteria and may contaminate the biopsy forceps and gastroduodenal tissue.

Conclusion

The tissue specimens from fundic gland polyp do not have the representative HP density for sampling. The prevalence of HP infection in human being should drop while most of the medical members have the concepts of HP impact. Suspicious histologic finding of HP density should add ancillary test such as $^{13}$C-urea Breath test or detection of HP antigen in stool for further confirmation.

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References


