

Research Article

When to Avoid Dialysis during Protein Purification?

Purushothaman Kavya^{1,3}, Sagar Krishna Bhat³, Shiva Siddappa¹, Rao AGA^{1,3}, Krishna Bhat Kadappu³ and Marathe GK^{1,2*}

¹Department of Studies in Biochemistry, University of Mysore, Manasagangothri, Mysuru, 570006, Karnataka, India

²Department of Studies in Molecular biology, University of Mysore, Manasagangothri, Mysuru, 570006, Karnataka, India

³Kaypeeyes Biotech Pvt Ltd, Hebbal Industrial Area, Mysuru 570016, Karnataka, India

Abstract

Membrane dialysis is a simple and widely used biochemical technique during protein purification to separate small molecular weight substances. In the process of isolating an industrially useful acid protease from locally available *Aspergillus* species, ammonium sulphate was used as salting-out agent in the very initial step of protein purification. While subjecting the ammonium sulphate precipitated-protease enriched sample for dialysis, we repeatedly found irregular perforations in the dialysis tubing. Careful analysis of the protease source suggested that, cellulase present in the protease enriched sample of *Aspergillus* species was responsible for this pore-forming act in the membrane. A simple gel filtration on Biogel-P100 (polyacrylamide beads) effectively separated ammonium sulphate from protease and cellulase, thereby avoiding dialysis. These studies provide one of the limitations of dialysis technique and suggest not to use cellulose-based dialysis tubing while dealing with samples containing cellulases.

Keywords: Ammonium Sulphate Precipitation; Cellulase; Dialysis; Protease

Introduction

Protein purification is vital to understand the structure and functions of any protein. The method of protein purification generally involves multiple chromatographic separation steps [1]. While purifying proteins from natural sources, concentrating the enzyme of our interest is a primary requisite. Commonly used concentrating

*Corresponding author: Marathe GK, Department of Studies in Biochemistry, University of Mysore, Manasagangothri, Mysuru, 570006, Karnataka, India, Tel: +91 9686423624; E-mail: marathe1962@gmail.com

Citation: Purushothaman K, Bhat SK, Siddappa S, Rao AGA, Kadappu KB et al., (2019) When to Avoid Dialysis during Protein Purification? Adv Ind Biotechnol 2: 009.

Received: March 21, 2019; **Accepted:** June 03, 2019; **Published:** June 10, 2019

Copyright: © 2019 Purushothaman K, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

methods include ultra filtration [2], reverse dialysis using sucrose [3], precipitation using chilled solvents (4°C to -20°C) [4,5] and salting out using divalent cationic salts [6]. Of these, salting out method of protein precipitation is commonly used. The most sought salt to perform salting out is ammonium sulphate [7]. However, it is essential to remove ammonium sulphate for subsequent purification of proteins, as it interferes with many of the further purification steps and protein quantification [8,9]. However, these conventional protein isolation methods are quite time consuming and involve multiple steps. Recently an Aqueous Two-Phase Flootation (ATPF) or Liquid Biphasic Flootation (LBF) technique has also been developed and successfully implemented in several instances [10,11].

Proteases derived from lower organisms have enormous industrial applications and researchers are on a quest for newer sources of proteases with high catalytic turnover and stability. In the process of searching for an industrially useful acid protease, we came across a strain of *Aspergillus niger* that produces thermostable acid protease. We initially enriched this enzyme using ammonium sulphate salting out method. The protease was salted out with 50% ammonium sulphate. To remove ammonium sulphate for the next step, we employed dialysis tubing with a molecular weight cut off 10KDa. We found dialysis tubing undergoing multiple, irregular perforations in the membrane, resulting in loss of the sample. We initially suspected a manufacturer's defect in dialysis tubing, but that was not the case. Upon careful analysis, we found that perforation was due to co-eluting cellulase(s) acting on dialysis tubing. We then successfully employed biogel-P100 (gel filtration matrix made out of polyacrylamide beads) to remove ammonium sulphate. We believe that these findings offer a precautionary message both to students and researchers alike while dealing with fungal enzymes.

Materials and Methods

Materials

Aspergillus niger culture was from Kaypeeyes biotech Pvt Ltd, Hebbal Industrial Area, Mysuru, Karnataka, India. Carboxymethyl-cellulose (CM-cellulose) (Sodium salt) and dialysis tubing (cellulose membrane) were obtained from SigmaAldrich, St. Louis, Missouri, USA; Haemoglobin was from MP Biomedicals, Santa Ana, California, USA; Biogel-P-100 was from Biorad, Hercules, California, USA; Spin columns were procured from GE healthcare, Little Chalfont, UK; Trichloroacetic Acid (TCA), Dinitrosalicylic Acid (DNS) and Nessler's reagent were obtained from Lobachemie, Mumbai, Maharashtra, India.

Methods

Production of acid protease from *A.niger*

A. niger was grown by solid state fermentation using wheat bran as substrate. To 100 g of wheat bran, 60 mL of 0.2 N HCl with trace minerals (70 mg each of CuSO₄, ZnSO₄ and FeSO₄ per 100 mL of 0.2NHCl) was added. Sterile wheat bran was inoculated with *A. niger* spore suspension and incubated at 30°C for 7 days. The extracellular

acid protease present in the fermented material was extracted in 0.1 M NaCl and recovered by centrifugation at 7000 g for 10 min at 4°C and filtered through Whatman filter paper no. 1. The clear supernatant was used as enzyme source.

Assay of acid protease

Acid protease was assayed as described by [4] with slight modifications. Briefly, 2% haemoglobin (acid denatured) prepared in 0.1 M acetate buffer pH 4.0 was used as substrate. To this, 0.4 mL (0.5-5µg) enzyme was added and incubated at 60°C for 10 min. The reaction was arrested by the addition of 2 mL of 5% TCA and the tubes were incubated at room temperature for 20 min. After 20 min, the reaction mixture was filtered through Whatman filter paper no.1 and the absorbance of the supernatant containing released tyrosine was measured at 280 nm. The amount of tyrosine released was calculated using a tyrosine standard (0-60 µg/mL). One unit of protease activity is defined as the amount of enzyme required to liberate 1µg of tyrosine per minute, under standard assay conditions.

Assay of cellulase

Cellulase assay was carried out as described by Wood and Bhat [12] with slight modifications. In brief, 0.1 mL (50-150µg) enzyme was added to 0.9 mL of 1% CM-cellulose prepared in 0.1 M acetate buffer pH 5.0. The enzyme reaction was carried out at 40°C for 120 min. The reaction was terminated by the addition of 1 mL of DNS reagent. The tubes were boiled in boiling water bath for 10 min, cooled and added with 8 mL of distilled water. The absorbance was read at 540 nm and the amount of glucose released was calculated using a glucose standard (0-1000 µg/mL). One unit of cellulase activity is defined as the amount of enzyme that liberates 1µmol of reducing sugar (expressed as glucose) per minute, under standard assay conditions.

Ammonium sulphate fractionation of acid protease

To the culture supernatant, ammonium sulphate was added to 30% saturation and then subsequently to 50% and 60%, with constant stirring at 4°C for 6 h. The precipitate was collected by centrifugation at 7000 g for 30 min at 4°C (in some experiments an aliquot from each fractionation was kept aside for assays of protease and cellulase).

Dialysis

50% ammonium sulphate pellet showing the highest specific activity was re-suspended in 50 mM acetate buffer pH 5.0 containing 0.1 M NaCl and dialyzed against same buffer.

Biogel-P100 chromatography

50% ammonium sulphate pellet showing the highest specific activity for protease was resuspended in 50 mM acetate buffer pH 5.0 containing 0.1 M NaCl. This sample (2ml containing 100mg protein) was loaded on to a 1.5cm × 78cm column packed with 137ml of Biogel P100 polyacrylamide beads, equilibrated with 50 mM acetate buffer pH 5.0 containing 0.1 M NaCl and eluted with the same buffer. Fractions were collected at a flow rate of 20ml/h and monitored for absorbance at 280nm. Each fraction was assessed for the presence of protease, cellulase and ammonia (see below).

Protein estimation

Protein estimation was carried out as described by and Biuret's method [13,14]. A) Lowry's method: This method involves two

reactions leading to color complex formation. In first reaction copper [II] reacts with the peptide bond in the protein under alkaline conditions, resulting in their reduction to cuprous [I] ions. The subsequent reaction involves the reduction of phosphomolybdicphosphotungstic acid to heteropolymolybdenum blue by the copper-catalyzed oxidation of aromatic acids. The blue complex formed is measured at 660 nm. The concentration of protein present in the sample is calculated using standard graph constructed with BSA as standard (15- 75 µg/mL). B) Biuret method: Under alkaline condition copper [II] ions reacts with peptide bond and forms mauve-colored coordination complex, which is measured at 540 nm. Amount of protein present is calculated using standard graph constructed with BSA (1-5 mg/mL).

Quantification of ammonia

Ammonia quantification was done using Nessler's reagent [15]. Each fraction from Biogel P-100 was treated with Nessler's reagent and absorbance was read at 425 nm. The amount of ammonia was calculated using a standard (1-20µg/ml) made with ammonia solution.

Results and Discussion

Dialysis is a simple technique, yet not useful for samples containing cellulases.

The global sales of industrial enzymes account for more than 3 billion USD of which proteases constitute more than 60% [16]. These proteases have wide range of applications and are mostly derived from microbial sources [17]. Among the microbes, enzymes derived from filamentous fungi such as *Aspergillus* species have many advantages over the other microbial sources, due to low cost of production, amenable for easy downstream processing, coupled with faster growth of the fungi and secretory nature of the enzyme [18]. While screening for an industrially useful protease from *Aspergillus* species, we came across an acid protease from *Aspergillus niger* which had industrially desirable qualities such as high catalytic turnover and thermostability. Hence, we decided to purify this fungal protease by conventional protein purification techniques. Precipitating the proteins by ammonium sulphate (salting out method) is a routine and simple technique widely used during protein purification. However, subsequent removal of residual ammonium sulphate is very much essential for further purification of the enzyme. In an effort to separate ammonium sulphate from the sample, dialysis was carried out using nitrocellulose membrane. But, every time during dialysis the sample was lost, as a result of formation of pores in the membrane (Figure 1). Although we suspected manufacture's defect initially, later we found an enzyme culprit responsible for this act. This culprit was cellulase. Accordingly, we found detectable cellulase activity in all the fractions that were also positive for proteases (Table 1). In order to separate ammonium sulphate we used spin columns (made of polyethersulfone) but this resulted in low recovery of the enzyme. Hence, we proceeded with differential ammonium sulphate fractionation, in an attempt to separate cellulase from protease. However, cellulase activity was also enriched with protease activity in all the steps of differential ammonium sulphate fractionation (Table 1). Therefore, we decided to use other chromatographic separation steps, which require prior removal of ammonium sulphate. Among the available methods, simple dialysis was the method of choice. However, we came across

the problem of cellulase action on dialysis tubing. Therefore, dialysis is not always the best technique for the removal of small molecules. Cellulases degrading cellulose is not new [19], but while purifying proteases co-purifying cellulases degrading cellulose was new to us. Similar difficulty was also encountered recently by Monico et al., where dialysis was not efficient in removing EDTA [20]. Other option for us at this stage was to use non-cellulosic dialysis membrane, but then they are expensive and often available in broad cut off range and not available with all the vendors. Hence, we decided to use biogel-P100.

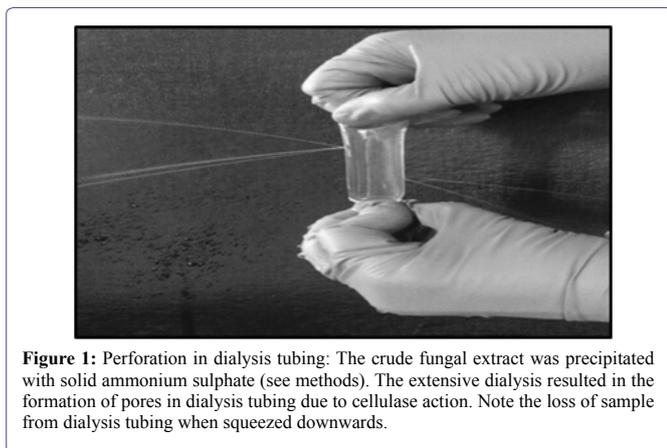


Figure 1: Perforation in dialysis tubing: The crude fungal extract was precipitated with solid ammonium sulphate (see methods). The extensive dialysis resulted in the formation of pores in dialysis tubing due to cellulase action. Note the loss of sample from dialysis tubing when squeezed downwards.

Biogel-P-100 effectively removes ammonium sulphate

Once we realized that cellulase was co-purifying with protease of our interest, carrying out dialysis using cellulose based membrane would be a futile exercise. Therefore, we attempted to remove ammonium sulphate by molecular sieving. A 50% ammonium sulphate pellet showed significant protease and cellulase activity (Table 1). This fraction was loaded on to a biogel P100 column. Loaded proteins were eluted (fraction number 20-42) in the first peak (Figures 2A and 2B), while the ammonium sulphate (fraction number 52-69) eluted later (Figures 2C and 2D). We confirmed elution of ammonium sulphate by both Nessler's reagent and measuring the conductance. The fractions containing proteins (fractions 20-42) showed no reactivity to Nessler's reagent while fractions containing ammonium sulphate reacted (fractions 52-69) to Nessler's reagent and showed conductance (Figures 2C and 2D).

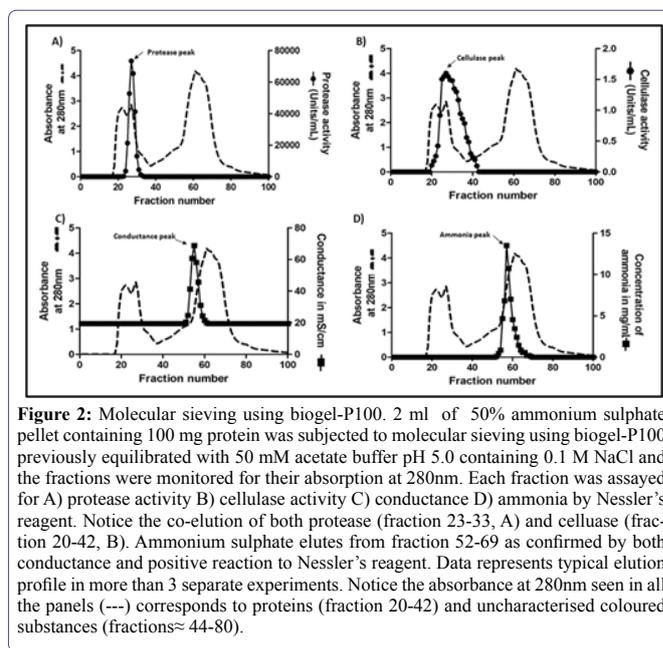


Figure 2: Molecular sieving using biogel-P100. 2 ml of 50% ammonium sulphate pellet containing 100 mg protein was subjected to molecular sieving using biogel-P100 previously equilibrated with 50 mM acetate buffer pH 5.0 containing 0.1 M NaCl and the fractions were monitored for their absorption at 280nm. Each fraction was assayed for A) protease activity B) cellulase activity C) conductance D) ammonia by Nessler's reagent. Notice the co-elution of both protease (fraction 23-33, A) and cellulase (fraction 20-42, B). Ammonium sulphate elutes from fraction 52-69 as confirmed by both conductance and positive reaction to Nessler's reagent. Data represents typical elution profile in more than 3 separate experiments. Notice the absorbance at 280nm seen in all the panels (---) corresponds to proteins (fraction 20-42) and uncharacterised coloured substances (fractions≈ 44-80).

Conclusion

Although dialyzing out ammonium sulphate is widely used during protein purification, it is not recommended specially for the samples containing cellulases. Molecular sieving using biogel-P100 instead of dialysis effectively removes ammonium sulphate from rest of the sample. As almost all fungi are known to have cellulases, along with other enzymes, commonly available dialysis membranes are likely to be degraded by the cellulase enzyme present in the fungal preparation. Hence, we highly recommend not using cellulose-based dialysis tubing while working with fungal enzymes.

Acknowledgement

Authors greatly acknowledge the financial support and *Aspergillus* culture from Kaypeeyes biotech Pvt. Ltd. Mysore, India. GKM thanks Institute of excellence, University of Mysore, UGC-Major Research Project, Vision Group of Science & Technology (VGST), Government of Karnataka and UGC-Special Assistance Program (UGC-SAP). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. Authors of this manuscript declare no conflict of interest.

Sample	Volume mL	Protein mg/ml	Protease		Cellulase	
			Activity U/mL	Specific activity U/mg	Activity U/mL	Specific activity U/mg
Crude extract	47	15.4	3835.8	249	2.64	0.171
Ammonium sulphate 30% supernatant	50	13	4319.9	332.2	1.49	0.114
Ammonium sulphate 30% pellet	4	50	4748.5	94.96	11.35	0.227
Ammonium sulphate 50% supernatant	49	8	95.2	11.9	-	-
Ammonium sulphate 50% pellet	4	53.8	37281.1	703.4	16.9	0.314
Ammonium sulphate 60% supernatant	46.5	6	12.87	6.4	-	-
Ammonium sulphate 60% pellet	0.8	16	5003.7	312.6	0.78	0.048

Table 1: Ammonium sulphate fractionation of protease. Calculated amount of ammonium sulphate was added to culture supernatant to achieve 30-60% saturation. Protease and cellulase activity were measured as described under methods. Both protease and cellulase co-precipitated at each saturation step and highest specific activity was seen in 50% pellet.

Author Contributions

Conceived and designed the experiments: GMK, AGA, Performed the experiments: KP, SKB, SS Analyzed the data: GMK, AGA Contributed reagents/materials/analysis tools: KBK, SKB, Wrote the paper: GMK, SS, KP.

References

1. Graslund S, Nordlund P, Weigelt J, Hallberg BM, Bray J, et al., (2008) Protein production and purification. *Nat methods* 5: 135-146.
2. Siala R, Sellami-kamoun A, Hajji M, Abid I, Gharsallah N, et al., (2009) Extracellular acid protease from *Aspergillus niger* II : purification and characterization. *Afr J Biotechnol* 8: 4582-4589.
3. Ire FS, Okolo BN, Moneke AA (2011) Purification and characterisation of an acid protease from *Aspergillus carbonarius*. *Afr J Food Sci* 5: 695-709.
4. Vishwanatha KS, Appu Rao AG, Singh SA (2009) Characterisation of acid protease expressed from *Aspergillus oryzae* MTCC 5341. *Food Chem* 14: 402-407.
5. Zanphorlin LM, Cabral H, Arantes E, Assis D, Juliano L, et al., (2011) Purification and characterization of a new alkaline serine protease from the thermophilic fungus *Myceliophthora* sp. *Process biochem* 46: 2137-2143.
6. Arakawa T, Timasheff SN (1984) Mechanism of protein salting in and salting out by divalent cation salts: Balance between hydration and salt binding. *Biochemistry* 23: 5912-5923.
7. Grodzki AC, Berenstein E (2010) Immunocytochemical methods and protocols-Antibody purification: Ammonium sulfate fractionation or gel filtration. *Methods mol biol* 588: 15-26.
8. Stanley PG (1963) Interference of ammonium sulfate with the estimation of protein by biuret reaction. *Nature* 197: 1108.
9. Niamke S, Kouame LP, Kouadio JP, Koffi D, Faulet BM, et al., (2005) Effect of some chemicals on the accuracy of protein estimation by the lowry method. *Biokemistri* 17: 73-81.
10. Lee SY, Khoiroh L, Ling TC, Show PL (2015) Aqueous two-phase flotation for the recovery of biomolecules. *Sep purif rev* 45: 81-92.
11. Sankaran R, Manickam S, Yap YJ, Ling TC, Chang JS, et al., (2018) Extraction of proteins from microalgae using integrated method of sugaring-out assisted Liquid Biphasic Flotation (LBF) and ultrasound. *Ultrason Sonochem* 48: 231-239.
12. Wood TM, Bhat TM (1998) Method for Measuring Cellulase Activities. In: *Cellulose and Hemicellulose, Methods enzymol*, New York academic press, USA, pg. no: 87-112.
13. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with Folin phenol reagent. *J Biol Chem* 193: 265-275.
14. Gornall AG, Bardawill CJ, David MM (1949) Determination of serum proteins by means of the biuret reaction. *J biol chem* 177: 751-766.
15. Bock JC (1941) Determination of urea nitrogen. *J Biol Chem* 140: 519-523.
16. Sumantha A, Larroche C, Pandey A (2006) Microbiology and Industrial Biotechnology of Food-Grade Proteases : A Perspective. *Food Technol Biotechnol* 44: 211-220.
17. De Souza PM, De Assis Bittencourt ML, Caprara CC, De Freitas M, De Almeida RPC, et al., (2015) A biotechnology perspective of fungal proteases. *Braz J microbiol* 46: 337-346.
18. Rao MB, Tanksale AM, Ghatge MS, Deshpande VV (1998) Molecular and biotechnological aspects of microbial proteases. *Microbiol mol biol rev* 62: 597-635.
19. Hurst PL, Nielsen J, Sullivan PA, Shepherd MG (1977) Purification and properties of a cellulase from *Aspergillus niger*. *Biochem J* 165: 33-41.
20. Monico A, Martinez Senra E, Canada FJ, Zorilla S, Perez Sala D (2017) Drawbacks of dialysis procedures for removal of EDTA. *PLoS One* 12: 1-9.



Journal of Anesthesia & Clinical Care
Journal of Addiction & Addictive Disorders
Advances in Microbiology Research
Advances in Industrial Biotechnology
Journal of Agronomy & Agricultural Science
Journal of AIDS Clinical Research & STDs
Journal of Alcoholism, Drug Abuse & Substance Dependence
Journal of Allergy Disorders & Therapy
Journal of Alternative, Complementary & Integrative Medicine
Journal of Alzheimer's & Neurodegenerative Diseases
Journal of Angiology & Vascular Surgery
Journal of Animal Research & Veterinary Science
Archives of Zoological Studies
Archives of Urology
Journal of Atmospheric & Earth-Sciences
Journal of Aquaculture & Fisheries
Journal of Biotech Research & Biochemistry
Journal of Brain & Neuroscience Research
Journal of Cancer Biology & Treatment
Journal of Cardiology: Study & Research
Journal of Cell Biology & Cell Metabolism
Journal of Clinical Dermatology & Therapy
Journal of Clinical Immunology & Immunotherapy
Journal of Clinical Studies & Medical Case Reports
Journal of Community Medicine & Public Health Care
Current Trends: Medical & Biological Engineering
Journal of Cytology & Tissue Biology
Journal of Dentistry: Oral Health & Cosmesis
Journal of Diabetes & Metabolic Disorders
Journal of Dairy Research & Technology
Journal of Emergency Medicine Trauma & Surgical Care
Journal of Environmental Science: Current Research
Journal of Food Science & Nutrition
Journal of Forensic, Legal & Investigative Sciences
Journal of Gastroenterology & Hepatology Research
Journal of Gerontology & Geriatric Medicine
Journal of Genetics & Genomic Sciences
Journal of Hematology, Blood Transfusion & Disorders
Journal of Human Endocrinology
Journal of Hospice & Palliative Medical Care
Journal of Internal Medicine & Primary Healthcare
Journal of Infectious & Non Infectious Diseases
Journal of Light & Laser: Current Trends
Journal of Modern Chemical Sciences
Journal of Medicine: Study & Research
Journal of Nanotechnology: Nanomedicine & Nanobiotechnology
Journal of Neonatology & Clinical Pediatrics
Journal of Nephrology & Renal Therapy
Journal of Non Invasive Vascular Investigation
Journal of Nuclear Medicine, Radiology & Radiation Therapy
Journal of Obesity & Weight Loss
Journal of Orthopedic Research & Physiotherapy
Journal of Otolaryngology, Head & Neck Surgery
Journal of Protein Research & Bioinformatics
Journal of Pathology Clinical & Medical Research
Journal of Pharmacology, Pharmaceutics & Pharmacovigilance
Journal of Physical Medicine, Rehabilitation & Disabilities
Journal of Plant Science: Current Research
Journal of Psychiatry, Depression & Anxiety
Journal of Pulmonary Medicine & Respiratory Research
Journal of Practical & Professional Nursing
Journal of Reproductive Medicine, Gynaecology & Obstetrics
Journal of Stem Cells Research, Development & Therapy
Journal of Surgery: Current Trends & Innovations
Journal of Toxicology: Current Research
Journal of Translational Science and Research
Trends in Anatomy & Physiology
Journal of Vaccines Research & Vaccination
Journal of Virology & Antivirals
Archives of Surgery and Surgical Education
Sports Medicine and Injury Care Journal
International Journal of Case Reports and Therapeutic Studies

Submit Your Manuscript: <http://www.heraldopenaccess.us/Online-Submission.php>