

Research Article

Long Term Remission of Hyper IgE Syndrome after Treatment with Cyclosporine-A: Clinic and Immunological Correlations

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Abstract

Hyper IgE syndrome is characterized by recurrent staphylococcal abscess, sinopulmonary infection, severe eczema and extremely high level of IgE. Recent work revealed a broader array of clinical features with defects in immune regulations. Management of these patients is very difficult because the pathophysiology of the immunodeficiency has not been completely elucidated. We examined the effect of a small dose of Cyclosporin-A (CSA) on the clinical course, and the excessive production of IgE and other immunologic parameters and infection in patients with Hyper IgE (HIE) syndrome.

Three patients, two females and one male, two were brother and sister; their ages were between 10 months and three years. All three patients were suffering from severe eczema, recurrent sinopulmonary infection, lung and skin abscesses, chronically draining ear and

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failure to thrive since the first few weeks of their lives. Their serum IgE was more than 10 times upper normal for age. Measurement of serum IgE, Cytokine IL-4, and IFN- γ and serum immunoglobulins measure before and after treatment, in addition, skin score of dermatitis and the number of infection, were evaluated before and after treatment with CSA.

Following treatment with CSA, 2-4 mg/kg per day in 2 divided doses, after standard treatment failed. There was a dramatic and sustained clinical improvement, especially dermatitis, associated with marked drop in serum IgE (P<0.0001), IL-4 (P<0.0001), and IFN- γ (P<0.001). There was no significant change in serum levels of IgG, IgA and IgM. The results of our study indicate that immune imbalance in HIE syndrome can be modulated by CSA that leads to marked drop on IgE and IL-4 synthesis and clinical remission. This treatment needs to be repeated in a larger number of patients.

Keywords: Cyclosporin-A; Cytokines; Dermatitis; Hyper IgE syndrome; Immunodeficiency; Interleukin-4

Introduction

The Hyper IgE (HIE) syndrome is a rare and complex disorder characterized by high serum IgE, chronic eczematoid dermatitis, recurrent sinopulmonary infections and skin abscesses [1]. The precise incidence is not known, with an estimated incidence ranging from 1 in 500,000 to 1 in 100,000 individuals. It is found equally in males and females, Caucasian, Asian and African [2,3]. The clinical features reported in 85 patients were as follows: skin abscesses 74%, eczema 58%, retained primary teeth 41%, fractures 39%, scoliosis 34%, and cancer 7% [4]. Recent studies have demonstrated that Peripheral Blood Mononuclear Cells (PBMCs) from HIE patients produce excessive IgE *in vitro*, spontaneously [5]. In addition, there is evidence that recombinant interleukin-4 (IL-4) induces IgE production in an unfractionated Peripheral Blood Mononuclear Cells (PBMC), which is enhanced by IL-2, IL-5 and IL-6 and suppressed by IFN- α , IFN- γ and PGE₂ [6]. However, no immediate causal relationship could be established between increased production of IgE and lymphokines secretion and the clinical course in HIE syndrome [7]. This leaves the etiology and pathogenesis of recurrent infections, increased serum IgE levels and eczematoid dermatitis in this syndrome poorly understood. However, recent studies showed that in the autosomal dominant and related disorders are due to defect in the Janus activated Kinase-signal transducer and activator of transcription (JAK-STAT) signaling pathway that is activated by cytokines, such as IL-6 and IL-2 [8].

Treatment of HIE patients is frustrating and remains largely unsuccessful. Several treatment modalities have been tried with limited and often temporary success [9-15]. Because HIE patients have excessive production of IgE and IL-4, clinical improvement may be achieved by down regulation of IgE and IL-4. Therefore, we conducted a prospective clinical immunological study to assess the effect of small doses of Cyclosporin-A (CSA) on the clinical course, especially dermatitis and skin infection in HIE syndrome. Three patients with the HIE syndrome showed remarkable clinical improvement following

treatment with a small dose of CSA 2-4 mg/kg per day. There was remarkable and significant drop in serum IgE, IL-4 and IFN- γ without affecting immunoglobulin levels after treatment compared to pre-treatment. Our study suggests that the clinical manifestations, especially dermatitis, may be related partially to the high levels of IL-4 and IgE levels which can be modulated by CSA. But these patients have many genetic defect in production of cytokines such as Th17 and IFN-gamma which impairs neutrophils, chemotaxis and IFN-gamma injections reduce the frequency of skin and lung infections [3,16]. Several immunological and pathological abnormalities have been observed in these patients including immunodeficiency markers such as elevated IgD levels, decreased IgG subclasses and poor response to both protein and polysaccharides immunization [16,17]. Some of these patients have symptoms of allergy to foods in 38% and anaphylaxis 8% [18]. These complex immunopathological abnormalities make management of these patients difficult.

Materials and Methods

Patients

Three children randomly selected who were diagnosed to have HIE syndrome according to the criteria defined by Buckley and Becker, were studied. There were two females and one male; two were a brother and sister [1]. Their ages ranged between 10 months and three years at the time of enrollment in the study. All patients had history of extensive dermatitis and recurrent staphylococcal abscesses of skin and lungs, sinusitis, and chronically draining ears since the first few weeks of life and they failed to thrive. They had serum IgE levels at least ten times the upper normal for age.

Scoring of skin inflammation

To assess the severity of dermatitis and the degree of inflammation prior to and during treatment with cyclosporin-A, a scoring system of 0-4 was used. 0 = no dermatitis or inflammation, 1 = mild dermatitis without infection on a small area of the skin, 2 = mild dermatitis involving less than 50% of the skin but no infection, 3 = extensive dermatitis involving most or all the skin but no infection or abscess formation, and 4 = the same as 3 but with superimposed infection and/or abscess formation.

Routine laboratory investigations

All patients had complete blood count and differential, hepatic and renal profile, surface and lesion cultures, chest and sinus X-ray at the time of admission and during the study period.

Treatment protocol

Patients were treated with appropriate antibiotics whenever indicated, antihistamines to control pruritus and topical and, at times, systemic steroids to control skin inflammation. After the above measures failed, patients were started on oral cyclosporin-A, at a dose of 2-4 mg/kg per day in two divided doses. Serum CSA was monitored to keep the trough level between 60 and 200 μ g/ml.

Serum IgE, IgG and IgA quantitation

Serum IgE was measured by Phadezym IgE PRIST using ELISA kit (Pharmacia, Uppsala, Sweden). IgG, IgM and IgA were measured by ACA Discrete Clinical Analyzer (Dupont, Wilmington, DE, USA).

Immunological studies

The following immunological investigations were carried out before and after CSA treatment i.e.,

- Immunophenotyping using Flow Cytometric Cell Analysis (FACS)
- Mitogenic response of PBMC to Phytohemagglutinin (PHA), Pokeweed Mitogen (PWM) and cytokine IL-2
- Natural Killer (NK) cell function
- Cytokine level of IL- α , IL-2, TNF- α , IFN- γ and IL-4

Isolation of PBMC from whole Blood

Whole peripheral blood from patients and normal healthy young individuals were collected into heparinized tubes. PBMCs were isolated by subjecting it to Ficoll-Hypaque gradient fractionation [19]. The isolated PBMC were washed twice with medium RPMI-1640 and finally suspended into complete medium in RPMI-1640, supplemented with Hepes 25 mM, L-glutamine 25 mM, 5% human AB serum, penicillin (100 U/ml), streptomycin (100 μ g/ml) and fungizone (20 μ g/ml) (Flow Laboratories, McLean, VA, USA). Cell counts and cell viability were judged by trypan blue dye (0.02%) exclusion method. PBMC with at least \geq 98% viability were used in the experiments.

Mitogens PHA and PWM Induced stimulation of PBMC

10^5 PBMC either from normal subjects or patients with Hyper IgE (HIE) syndrome, were cultured in quadruplicate in 96 well round bottom microtiter plate in a final volume of 250 μ l complete RPMI-1640 containing 5 μ g/10⁶ml of PHA or PWM (Flow Lab, McLean, VA, USA) for 72 hours. The proliferation response was measured from the [³H] Thymidine [³HTdR] uptake after an 18 hour pulse.

(1 μ Ci/well Sp. activity 6.7 μ Ci/mM) as given by the β counts of cultures harvested with a cell harvester (Titretek Flow Labs, Rockville, MD) [20].

Interleukin-2 dependent proliferation of PBMC

As previously described, 10^5 PBMC either from normal individuals or hyper IgE patients were cultured in the presence of 100 U of recombinant IL-2 (Genzyme, Boston, MA, USA) for 72 hours. The proliferative response, ³HTdR uptake was then measured [21].

Natural Killer (NK) cell assay

This was performed using a standard 4 hour ⁵¹Cr release assay as previously described [22]. K562 erythroleukemia (NK sensitive) cells were labeled with Sodium Chromate and used as target for normal or patients PBMC (effector cells) at various 12:5:1 to 100:1, effector:target ratio.

The percent specific cytotoxicity was computed as follows:

$$\% \text{ Specific Cytotoxicity} = \frac{{}^{51}\text{Cr CPM Experimental release} - {}^{51}\text{Cr CPM Spontaneous release}}{{}^{51}\text{Cr CPM Maximum release} - {}^{51}\text{Cr CPM Spontaneous release}} \times 100$$

Quantitations of cytokines

As earlier described Interleukin-1 α (IL-1 α), Interleukin-2 (IL-2), Interferon- γ (IFN- γ), Tumour Necrosis Factor- α (TNF- α) and Interleukin-4 (IL-4) were measured from the plasma samples of HIE patients and normal individuals using highly sensitive ELISA kits from

Endogen MA, USA, (IL- α , TNF- α), Genzyme MA, USA, (IL-2), Holland Biotechnology (IFN- γ) and R&D System MN, USA, (IL-4). Manufacturer's guidelines were followed in carrying out the measurements of these cytokines [23].

Flow Cytometric (FACS) analysis

Heparinized blood was used to quantify various populations and subsets of lymphocytes using fluorescent-phycoerythrin conjugated monoclonal antibodies (FITC/PE-MOAbs Becton-Dickinson CA, USA). Cells were labeled with FITC or PE-MOAbs Leu⁴/12, Leu³/2, Leu⁴/DR, Leu⁷/2, Leu³/8, Leu⁴/11+19 for 30 minutes at 4°C, RBC were lysed and cells were fixed in PBS containing 1% paraformaldehyde. Percent positive cells for various surface markers were analyzed using FACS Scan (Becton-Dickinson CA, USA) [22].

Statistical evaluation

The results were expressed as mean \pm SE. Statistically, significance of difference between various experimental groups was tested using one way analysis of variance and students t test. p value at ≤ 0.05 was considered significance.

Results

All three patients met the criteria of HIE diagnosis. Their serum IgE levels ranged between 5,400 and 38,400 (normal 3.84-72 μ g/L) at the time of diagnosis and before starting treatment with CSA. All patients required repeated hospitalization for treatment of infections and/or surgical drainage of skin abscesses. One patient had keratoconjunctivitis and lost one eye of pseudomonas infection. The other two had mucocutaneous candidiasis. They also had chronic sinusitis and bronchiectasis. *Staphylococcus aureus* was cultured from all lesions. Their peripheral blood count showed significant eosinophilia (23-65%). Specific IgE antibody measurement *in vitro* by RAST scores were 3-4 against milk, eggs and wheat. None of the patients experienced respiratory allergies, although one patient had wheezing episodes during lung infections.

Patients No 1 and 2 were sister and brother with negative family history of a similar condition. Patient No 3 had 6-year-old maternal cousin who died with identical history. The parents of all our patients (six other patients not reported here) were of consanguine marriage, usually first degree cousins (Figure 1).

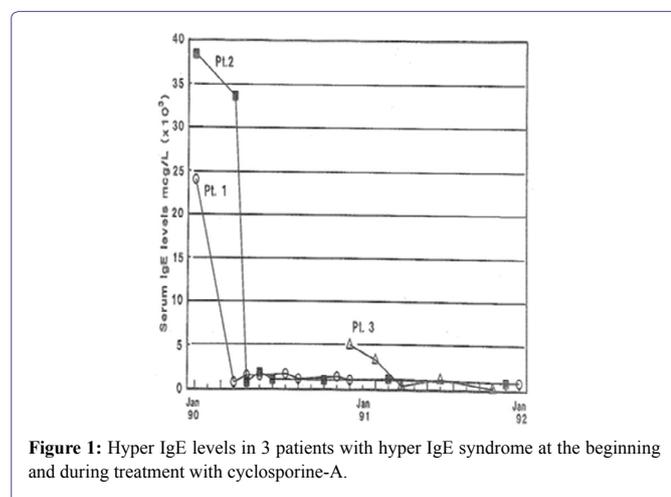


Figure 1: Hyper IgE levels in 3 patients with hyper IgE syndrome at the beginning and during treatment with cyclosporine-A.

Response to Treatment

On admission the skin score was 4 in all patients. There was little or insignificant response of antihistamines, skin emollients, steroids and repeated courses of antibiotics. There was a dramatic and a marked improvement within 72 hours of starting CSA treatment. Inflammation cleared up from most of the skin; skin scores dropped to 1-2 within 3 to 7 days after starting CSA. This improvement was constant and continued throughout the treatment course. Twenty-four months after starting CSA patients 1 & 2 and after 16 months, patient 3 continued to do well with minimal respiratory infections. This improvement coincided with a sharp and a significant drop in serum IgE levels. Serum IgG, IgA and IgM were not affected except patient no 1 where the posttreatment IgG level increased from 3 grams to 28 gram/L. In patient no 1 serum IgE level dropped from 24,000 μ g to 800 μ g/L in 2 weeks, patient no 2 serum IgE dropped from 38,400 μ g to 864 μ g/L in 7 weeks, while patient no 3 serum IgE level dropped from 5,040 μ g to 480 μ g/L in 10 weeks. Although serum IgE levels stayed above the normal range, none of the patients had levels near the pre-treatment values (Table 1).

Proliferative response (³HTdR uptake) to mitogens and cytokine

PBMC from the hyper IgE patients showed impaired response to PHA and PWM as compared to control individuals with (3-37%) suppression for PHA, and 4-43% suppression for PWM. Similar suppression (8%-22%) was also noted when PBMC from hyper IgE patients were co-cultured with interleukin-2 (Table 2).

Analysis of Natural Killer (NK) cell function

The results of NK assay performed with fresh PBMC of patients even at higher effector: target ratio (100:1) failed to demonstrate any appreciable level of cytotoxicity to K562 tumor target cells as compared to normal PBMC controls (Figure 2).

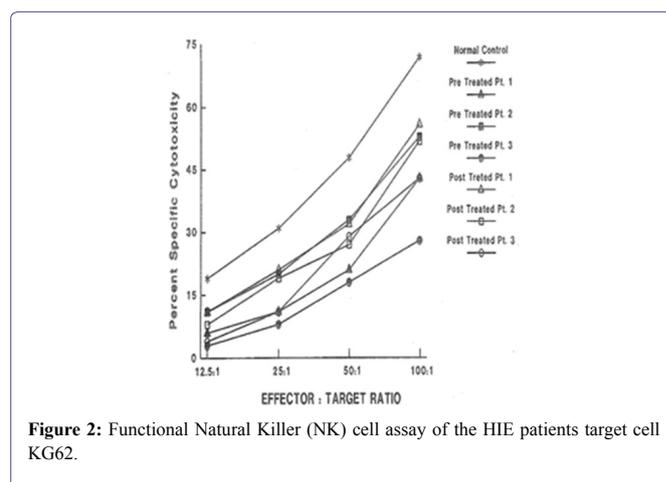


Figure 2: Functional Natural Killer (NK) cell assay of the HIE patients target cell KG62.

Plasma level of cytokines

The quantitation of cytokines prior to cyclosporin-A treatment indicated elevated level of IL-4 (112-147 units/ml), IFN- γ (11-56 U/ml). Plasma level of IL-2 (results not presented), IL-1 α and TNF- α were either very low or not detected in any of the hyper IgE patients. A highly significant ($P < 0.0001$) lower levels of IL-4 was observed in all post treated patients (Figures 3a and 3b).

Patient	Age	Sex	Igs Normal Range (gm/l)	Serum (µg/ml)			
				IgG (4.4-12.9)	IgA (0.4-1.2)	IlgM (0.2-1.7)	IgE (3.84-72)
1	2 years	F	Pre-tr.	3.0	0.6	0.4	24.000 ^a
			Post-tr.	28.2	1.4	0.5	1.104 ^a
2	10 months	M	Pre-tr.	5.3	0.4	0.8	38.400 ^a
			Post-tr.	13.3	1.3	1.4	888 ^a
3	3 years	F	Pre-tr.	13.2	2.6	1.8	5.040 ^a
			Post-tr.	13.3	ND	1.2	211 ^a

Table 1: Serum IgE levels in 3 patients with HIE syndrome.

A highly significant difference ($P < 0.0001$) in IgE plasma level was observed in all the 3 HIE-patients after CSA treatment. No statistical significant difference was observed in the level of IgM and IgA. Whereas significant difference ($P < 0.002$ Pt.1) was observed in IgG level.

Patient		PHAa	PWMa	IL-2b
1	Pre-treatment	42453±2363	28112±1983	49008±3097
	Post-treatment	63017±2311	50307±4763	60946±1781
2	Pre-treatment	54583±1932	43895±1108	54034±3781
	Post-treatment	64961±1851	46388±3192	59689±1673
3	Pre-treatment	55257±2117	47348±3013	54034±1907
	Post-treatment	59387±3201	46407±2011	59921±2307
	Normal Control (n=10)	67387±3217	49321±2703	62831±3781

Table 1: HIE Patients PBL proliferative response [3HTdR CPM uptake] to mitogen or cytokine.

^aPHA and PWM were used at 5 µg/1x50⁶ PBL/ml concentration.

^bInterleukin-2 was used at 100 u/1x10⁶/PBL/ml. 105PBL/well/250µg were cultured for 72 hours at 37°, 5% CO₂ and pulsed with 1 µci3HTdR for 18 hours before harvesting.

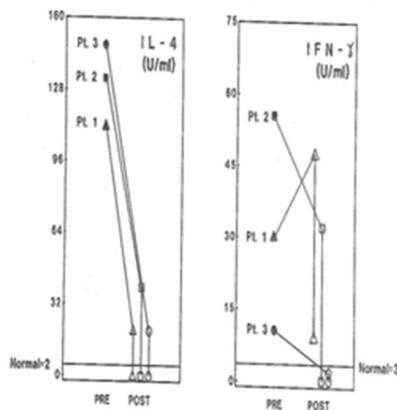


Figure 3a: Plasma level of cytokine IL-4 and IFN- γ before and after treatment with CSA in 3 patients with HIE syndrome.

Flow cytometry analysis

The absolute numbers of T and B lymphocytes did not demonstrate any significant changes other than that of normal controls. Whereas the number of T helper cells in all patients was substantially less as compared to normal individuals at the start of the treatment. The T4 (helper T cell): T8 (suppressors T cell) ratio was significantly

($P < 0.05$) less than normal Saudi controls. The T4:T8 ratio showed a significant increase after CSA treatment reflecting increase in T helper and decrease in T suppressor cell population. A higher degree of activated T cells (Leu4 /DR) were also found in all the patients (Table 3).

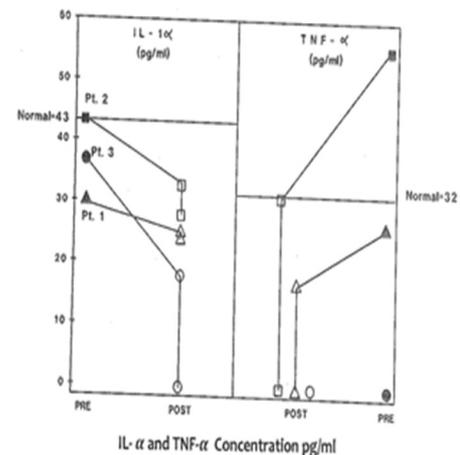


Figure 3b: Plasma level of cytokine IL- α and TNF- α before and after treatment with CSA in 3 patients with HIE syndrome.

Patients	Percent Positive cells for various sub-populations of lymphocytes							
		T	B	T4	T8	Ratio	Act.T	Nk
1	Pre-treatment	70	18	17	44	0.3a	56b	5c
	Post-treatment	73	26	23	39	0.5	43	7
2	Pre-treatment	68	18	10	57	0.2 ^a	40 ^b	5 ^c
	Post-treatment	63	27	42	38	1.1	33	8
3	Pre-treatment	50	17	24	31	0.8a	13	2c
	Post-treatment	66	27	36	24	1.5	11	7
Normal Control		78±7	16±7	39±9.1	35±8.7	1.1(p<0.05) ^a	11±4(p<0.002) ^b	16±3(p<0.02) ^c

Discussion

Our current knowledge about the pathophysiology, pathogenesis, and definite treatment of HIE Syndrome (JOB's) is limited. A major disbalance and altered immune responses are implicated in HIE patients. Some of the immune disorders are due to defects in the activator of transcription (JAK-STAT) signaling pathway in the autosomal dominant Hyper-IgE, which is activated by interleukin IL-6 and IL-2. This defect leads to impaired T cell helper type Th17 [3]. Though at present the immunosuppressant drug CSA has been widely used to prevent allograft rejection its successful use to treat variety of skin diseases particularly psoriasis and severe atopic eczema has also been reported [24-31]. CSA is a potent inhibitor of T cell activation and subsequent production of various immune mediators namely IL-2, IL-4 and IFN- γ [32-35]. IL-4, IL-5 and IL-13 have been shown to mediate IgE synthesis in human involving a complex interaction among B cells, T cells and monocytes [36-39]. The whole cascade of IgE production can be modulated positively or negatively by cytokines. To elucidate the underlying immunological mechanism in HIE patients, we have presented the results of immunophenotyping, proliferative response of PBMC to PHA, PWM and IL-2, cytotoxicity level of NK cells, levels of various cytokines and immunoglobulins before and after low dose if CSA treatment of three HIE patients.

In response to low dose (60 - 200 μ g/ml Plasma level) CSA treatment, the results of immunoglobulins quantitation showed a remarkable drop in the IgE level in all three patients without any adverse effects of IgM, IgG and IgA immunoglobulins. It is a well accepted phenomenon in mice that subsets of helper T cells, Th1 and Th2, are responsible for differential production of cytokines [40]. Th1 clone when stimulated with appropriate stimulus can produce IL-2, IFN- γ and TNF- α ; whereas Th2 clone produces IL-4, IL-5, IL-6, and IL-10. IFN- γ has been shown to be a very important mediator in curtailing the production of IgE, whereas IL-4 molecules support the primary induction and production of IgE [6,41-43]. A similar dichotomy among human T lymphocyte function has been observed *in vitro* [44]. Our results of flow cytometric analysis are supportive of this notion since absolute number of T and B lymphocytes in HIE patients were not altered, but rather a low T4:T8 ratio was observed in the pre-treated patients. The results of proliferative response to PHA, PWM, and IL-2 indicated inherent defect in either T cell or T cells bearing receptor for IL-2 in HIE patients. The NK cell dependent cytotoxicity in all the three pre-treated patients was substantially low as compared to normal control. An earlier study by Wehrmann et al., demonstrated the number and function of NK cells were altered in atopic patients with high level of circulating IgE [45]. The results of our *in vitro* study have shown that IgE at higher concentration can decrease

regulative killer lineage cells function, implicating similar role of IgE in HIE patients [46]. Our results of impaired PHA response are in disagreement with earlier work. However the response of PWM in HIE patients has been controversial, we found mild to severe suppression of PWM proliferative response prior to CSA treatment with a tendency to improve with CSA treatment [13]. The results of cytokines measurement in our study indicate that IL-1 α , TNF- α and IL-2 do not play any significant role in the pathophysiology of HIE Syndrome. Recent reports by Mogensen and Ochs et al., found that susceptibility to infection is related in part to defect in Th17 which results in decreased neutrophil proliferation and chemotaxis decreased inflammation and susceptibility to Candida and bacteria infection [3,47]. The earlier *in vitro* findings using PMA to stimulate PBMC from HIE patients also showed that inducible IL-2 production is least affected in HIE patients [7]. Our findings of high level of IL-4 and IgE in HIE patients before treatment are confirmatory of previous studies [7]. The most striking findings of this study, after CSA treatment, are that levels of IgE and IL-4 dropped to normal control levels. Following CSA treatment, a sharp decrease to an undetectable level of IFN- γ was observed (P>0.001).

Previous treatment of HIE patients with a variety of agents and immune modulators, including levamisole, cimetidine, transfer factor and ascorbic acid, isotretinoin, cromoglycate and repeated plasma pheresis, failed to induce long term clinical remissions in these patients [9-15,48]. Although interferon- γ was shown *in vitro* to regulate the IgE production by PBMC from HIE patients, it had no significant clinical effect in treating these patients [41].

Whether this clinical improvement and the normalization of the immune imbalance in HIE patients following treatment with CSA is long lasting and void of serious complication, needs to be evaluated further. After two years of treatment, there is no evidence of any complications except mild hypertrichosis in one patient. On one occasion, the skin lesion flared up three days after CSA was stopped suggesting, at least, that the skin improvement is directly related to CSA therapy.

Recently, low dose cyclosporin-A has been used to treat cases of severe atopic dermatitis with no subsequent serious complications [31,49,50]. Similarly safe and beneficial results after treatment with CSA in a variety of dermatological disorders have been reported [51]. The response of patients with HIE syndrome and severe atopic dermatitis to low dose CSA suggests a common link between these disorders. Both have immune dysregulation, with over production of IgE, severe pruritus and recurrent skin infection with staph aureus. This link needs further investigations in order to uncover possible common mechanism for causing the syndrome.

Though much work is to be done to elucidate the precise mechanism of beneficial effect of CSA treatment in HIE patients, the results discussed here are encouraging for the future use of CSA to treat HIE patients. The effects of a newly and highly immunosuppressive agent FK506 on the IL-4 induced IgE production is also being examined *in vitro* to find out if it has similar effect as of CSA.

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References

1. Buckley RH, and Becker WG (1978) Abnormalities in the regulation of human IgE synthesis. *Immunol Rev* 41: 288-313.
2. Woeliner Woellner C, Gertz EM, Schäffer AA, Lagos M, Perro M, et al. Mutations in STAT3 and diagnostic guidelines for hyper-IgE syndrome. *J Allergy Clin Immunol* 125: 424-432.
3. Mogensen TH (2013) STAT3 and the Hyper-IgE syndrome: Clinical presentation, genetic origin, pathogenesis, novel findings and remaining uncertainties. *JAKSTAT* 2: 23435.
4. Gernez Y, Freeman AF, Holland SM, Garabedian E, Patel NC, et al. (2018) Autosomal Dominant Hyper-IgE Syndrome in the USIDNET Registry *J Allergy Clin Immunol Pract* 6: 996-1001.
5. Leung DY, Geha RS (1988) Clinical and immunologic aspects of the hyperimmunoglobulin E syndrome. *Hematol Oncol Clin North Am* 2: 81-100.
6. Pene J (1989) Regulatory Role of Cytokines and DC23 in the Human IgE antibody synthesis. *Int Arch Allergy Immunol* 90: 32-40.
7. Vercelli D, Jabara HH, Cunningham-Rundles C, Abrams JS, Lewis DB, et al. (1990) Regulation of Immunoglobulin (Ig)E Synthesis in the Hyper-IgE Syndrome. *J Clin Invest* 85: 1666-1671.
8. Alsum Z, Hawwari A, Alsmadi O, Al-Hissi S, Borrero E, et al. (2013) Clinical, immunological and molecular characterization of DOCK8 and DOCK8-like deficient patients: single center experience of twenty-five patients. *J Clin Immunol* 33: 55-67.
9. Businco L, Laurenti F, Rossi P, Galli E, Aiuti F (1981) A child with atopic features raise serum IgE, and recurrent infection treated with levamisole. *Arch Dis Child* 56: 60-63.
10. Friedenbergr WR, Marx JJ Jr, Hansen RL, Haselby RC (1979) Hyperimmunoglobulin E Syndrome: Response to transfer factor and ascorbic acid therapy. *Clin Immunol Immunopathol* 12: 132-136.
11. Mawhinney H, Killen M, Fleming WA, Roy AD (1980) The hyperimmunoglobulin E syndrome--A neutrophil chemotactic defect reversible by histamine H2 receptor blockade? *Clin Immunol Immunopathol* 17: 483-491.
12. Leung DYM, Wood NL, Geha RS (1985) Reversal of cellular abnormalities in the hyper IgE syndrome following plasmapheresis. *Clin Res* 33: 161A.
13. Geha RS, Leung DYM (1989) Hyper Immunoglobulin E Syndrome. *Immunodef. Rev* 1: 155-172.
14. Shuttleworth D, Holt PJA, Mathews N (1988) Hyper Immunoglobulin E syndrome: treatment with isotretinoin. *Br J Dermatol* 119: 93-99.
15. Yokota S, Mitsuda T, Shimizu H, Ibe M, Matsuyama S (1990) Cromoglycate treatment of patient with hyperimmunoglobulinaemia E syndrome. *Lancet* 335: 857-858.
16. Buckley RH (2001) The hyper-IgE syndrome. *Clin Rev Allergy Immunol* 20: 139-154.
17. Sheerin KA, Buckley RH (1991) Antibody responses to protein, polysaccharide, and phi X174 antigens in the hyperimmunoglobulinemia E (hyper-IgE) syndrome. *J Allergy Clin Immunol* 87: 803-811.
18. Siegel AM, Stone KD, Cruse G, Lawrence MG, Olivera A, et al. (2013) Diminished allergic disease in patients with STAT3 mutations reveals a role for STAT3 signaling in mast cell degranulation. *J Allergy Clin Immunol* 132: 1388-1396.
19. Boyum A (1968) Isolation of mononuclear cells and granulocytes by one step centrifugation and of granulocytes by combining centrifugation and sedimentation at Ig. *Scand J Clin Lab Invest* 97: 77-98.
20. Kwaasi AA, Parhar RS, Harfi H, Tipirneni P, al-Sedairy ST (1992) Characterization of antigens and allergens of date palm (*Phoenix dactylifera*) pollen. Immunological assessment of atopic patients using whole extract or it fraction. *Allergy* 47: 535-44.
21. Parhar RS, Yagel S, Lala PK (1989) PGE2-mediated immunosuppression by first trimester human decidual cells blocks activation of maternal leukocytes in the decidua with potential anti-trophoblast activity. *Cell Immunol* 120: 61-74.
22. Parhar RS, Ernst P, Sheth KV, al-Sedairy S (1992) Anti-Tumor cytotoxic potential and effect on human bone marrow GM-CFU of human LAK cells generated in response to various cytokines. *Eur Cytokine Netw* 3: 299-306.
23. Bouchama A, Parhar RS, el-Yazigi A, Sheth K, al-Sedairy S (1991) Endotoxemia and release of Tumournecrosis factor and interleukin 1 alpha in acute heatstroke. *J Appl Physiol* 70: 2640-2644.
24. Borel JF, Feurer C, Magñee C, Stähelin H (1977) Effects of the new anti-lymphocytic peptide cyclosporin A in animals. *Immunology* 32: 1017-1025.
25. Calne RY, White DJ, Evans DB, Thiru S, Henderson RG (1981) Cyclosporin A in cadaveric organ transplantation. *Br Med J (Clin Res Ed)* 282: 934-936.
26. Calne RY, Rolles K, White DJ, Thiru S, Evans DB, et al. (1979) Cyclosporin A initially as the only immunosuppressant in 34 recipients of cadaveric organs: 32 kidneys, 2 pancreases, and 2 livers. *Lancet* 2: 1033-1036.
27. Starzl TE, Klintmalm GB, Porter KA, Iwatsuki S, Schröter GP (1981) Liver transplantation with use of cyclosporin a and prednisone. *N Engl J Med* 305: 266-269.
28. Kahan BD (1989) Cyclosporine. *The New England Journal of Medicine* 321: 1725-1738.
29. Biren CA, Barr RJ (1986) Dermatologic applications of cyclosporine. *Arch Dermatol* 122: 1028-1032.
30. Griffiths CE, Powles AV, Leonard JN, Fry L, Baker BS, et al. (1986) Clearance of psoriasis with low dose cyclosporin. *Br Med J (Clin Res Ed)* 293: 731-732.
31. Van Joost T, Stolz E, Meule F (1987) Efficacy of low dose cyclosporine in severe atopic skin disease. *Arch Dermatol* 123: 166-167.
32. Britton S, Palacios R (1982) Cyclosporin A-Useful, risks and mechanism of action. *Immunol Rev* 65: 5-22.
33. Bunjes D, Hardt C, Röllinghoff M, Wagner H (1981) Cyclosporin A mediates immunosuppression of primary cytotoxic T cell responses by impairing the release of interleukin 1 and interleukin 2. *Eur J Immunol* 11: 657-661.
34. Reem GH, Cook LA, Vilcek J (1983) Gamma interferon synthesis by human thymocytes and T lymphocytes inhibited by cyclosporin A. *Science* 221: 63-65.
35. Wang SC, Zeevi A, Jordan ML, Simmons RL, Twardy DJ (1991) FK 506, rapamycin, and cyclosporine: effects on IL-4 and IL-10 mRNA levels in a T-helper 2 cell line. *Transplant Proc* 23: 2920-2922.

36. DeKruiff RH, Turner T, Abrams JS, Palladine MA, Umetsu DT (1989) Induction of human IgE synthesis by CD4+ T cell clones. Requirement for interleukin 4 and low molecular weight B cell growth factor. *J Exp Med* 170: 1477-1494.
37. Ishizaka A, Sakiyama Y, Nakanishi M, Tomizawa K, Oshika E, et al. (1990) The inductive effect of interleukin-4 on IgG4 and IgE synthesis in human peripheral blood lymphocytes. *Clin Exp Immunol* 79: 392-396.
38. Rousset F, Robert J, Andary M, Bonnin JP, Souillet G, et al. (1991) Shifts in interleukin-4 and interferon-gamma production by T cells of patients with elevated serum IgE levels and the modulatory effects of these lymphokines on spontaneous IgE synthesis. *J Allergy Clin Immunol* 87: 58-69.
39. Romagnani S, Maggi E, Del Prete G, Parronchi P, Tiri A, et al. (1989) Role of interleukins in induction and regulation of human IgE. *Clin Exp Rheumatol* 7: 117-122.
40. Coffman RL, Seymour BW, Lebman DA, Hiraki DD, Christiansen JA, et al. (1988) The role of helper T cell products in mouse B cell differentiation and isotype regulation. *Immunol Rev* 102: 5-28.
41. King CL, Gallin JI, Malech HL, Abramson SL, Nutman TB (1989) Regulation of immunoglobulin production in hyperimmunoglobulin E recurrent-infection syndrome by interferon gamma. *Proc Natl Acad Sci USA* 86: 10085-10089.
42. Ishizaka A, Joh K, Shibata R, Wagatsuma Y, Nakanishi M, et al. (1990) Regulation of IgE and IgG4 synthesis in patients with hyper IgE syndrome. *Immunology* 70: 414-416.
43. Gauchat JF, Lebman DA, Coffman RL, Gascan H, de Vries JE (1990) Structure and expression of germline epsilon transcripts in human B cells induced by interleukin 4 to switch to IgE production. *J Exp Med* 172: 463-473.
44. Del Prete G, Maggi E, Parronchi P, Chrétien I, Tiri A, et al. (1988) IL-4 is an essential factor for the IgE synthesis induced *in vitro* by human T cell clones and their J *Immunol* 140: 4193-4198.
45. Wehrmann W, Reinhold U, Kuekl S, Franke N, Uerlich M, et al. (1990) Selective Alteration in natural killer cell subsets in patients with atopic dermatitis. *Int Arch Allergy Appl. Immunol* 92: 318-322.
46. Parhar RS, Ernst P, Al-Mohanna F, Kwaasi A, Sheth KV, et al. (1991) 425 *In vitro* regulation of immune cell functions by human IgE. *J Allergy and Clinical Immunol* 87: 2-246.
47. Ochs HD, Oukka M, Torgerson TR (2009) TH17 cells and regulatory T cells in primary immunodeficiency diseases. *J Allergy Clin Immunol* 123: 977-983.
48. Donabedian H, Alling DW, Gallin JI (1982) Levamisole is inferior to placebo in the hyperimmunoglobulin E recurrent-infection (Job's) syndrome. *N Engl J Med* 307: 290-292.
49. Logan RA, Camp RD (1988) Severe atopic eczema: response to oral cyclosporin A. *J R Soc Med* 81: 417-418.
50. Taylor RS 3rd, Cooper KD, Headingto JT, Ho VC, Ellis CN, et al. (1989) Cyclosporine therapy for severe atopic dermatitis. *J Am Acad Dermatol* 21: 580-583.
51. Ho VC, Lui H, McLean DI (1990) Cyclosporine in nonpsoriatic dermatoses. *J Am Acad Dermatol* 23: 1248-1259.



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