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Research Article

Application of Ginsenoside-Rg1 Enhanced Glutathione Peroxidase Activity but could not Ameliorate Hyperoxia-Induced Vascular Endothelial Oxidative Stress and Apoptosis

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Abstract

Objective: Oxygen therapy is necessary to preterm infants with respiratory distress. However, hyperoxia may cause bronchopulmonary dysplasia and retinopathy of prematurity due to suppression of vasogenesis and increase of cell death. Ginsenoside-Rg1, one of the active components of Ginseng, is shown as a proangiogenic fac-

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tor of vascular endothelial cells. We evaluated whether application of ginsenoside-Rg1 is able to improve hyperoxic-induced vascular endothelium injury.

Methods: Human umbilical vein endothelial cells (HUVECs) were cultured under room air and 60% oxygen for 72 hours, respectively. Gensenoside-Rg1 was added to the medium at 0, 75, 150, 300nM. HUVECs proliferation, oxidative stress and apoptosis under normoxic and hyperoxic conditions were assayed by Western Blot.

Results: Under hyperoxia (60% O2), HUVECs proliferation and levels of vascular endothelial growth factor (VEGF) were significantly decreased after ginsenoside-Rg1 treatment. However, both levels of glucocorticoid receptor (GR) and glutathione peroxidase (GPx) were increased after 72 hours ginsenoside-Rg1 treatment, but no changed under room air control. The levels of oxidative stress-induced Bax and cytochrome c and apoptosis-related active caspase 3 and poly ADP-ribose polymerase were significantly increased after ginsenoside-Rg1 treatment under hyperoxic condition.

Conclusion: In HUVECs model, ginsenoside-Rg1 is unable to overcome the major hyperoxic-induced vascular endothelium injury. It might aggravate oxidative stress and endothelial apoptosis caused by oxygen toxicity. However, both elevated levels of GR and GPx indicate that ginsenoside-Rg1 could be involved in vascular signaling and the regulation of oxidative stress under hyperoxia. Further investigation of Rg1 effects under hyperoxia is required.

Keywords: Apoptosis; Ginsenoside-Rg1; Hyperoxic Injury; Oxidative Stress; Vascular Endothelial Cel

Abbreviations

BPD: Bronchopulmonary Dysplasia

Cyt c: Cytochrome c

ECGS: Endothelial Cell Growth Supplement

eNOS: Endothelial Nitric Oxide Synthease

FBS: Fetal Bovine Serum

GPx: Glutathione Peroxidase

GR: Glucocorticoid Receptor

HUVECs: Human Umbilical Vein Endothelial Cells

PARP: Poly ADP-Ribose Polymerase

ROP: Retinopathy of Prematurity

ROS: Reactive Oxygen Species

VEGF: Vascular Endothelial Growth Factor

Introduction

Oxygen administration is necessary to preterm infants with respiratory stress, but hyperoxia is shown to be correlated with several

diseases in preterm infants [1], such as bronchopulmonary dysplasia (BPD) and Retinopathy of Prematurity (ROP) [2]. Under hyperoxia, the characteristic of injury includes inflammatory cell influx, endothelial and epithelial cell death, and vasogenesis suppression [3]. Vascular endothelial cells appear to be one of the major targets of hyperoxic injury. From the pathophysiology, hyperoxia disrupts the survival and functions of vascular endothelial cells, which contributes to the development of diseases [4]. Hyperoxia also alters Vascular Endothelial Growth Factor (VEGF) signaling pathway in both BPD and ROP [1], and high concentration oxygen administration releases Reactive Oxygen Species (ROS), which is believed to cause oxidative stress. Moreover, hyperoxic microenvironment is associated with multiple alternations in the extracellular and intracellular milieu of vascular endothelium that oxidative stress injury may act as inducers of apoptosis and cell death.

Ginseng, the root of Panax ginsengg, has been used for centuries as a component of traditional complementary medicine [5]. It could be used as a tonic agent to combat stress, or a medicine to improve cardio-pulmonary function. The molecular compositions of ginseng have been studied extensively. Ginsenosides are the constituents that responsible for the actions of ginseng, and Rg1 is among the most abundant and active components. Recent *in vitro* studies showed that ginsenoside-Rg1 is a proangiogenic factor of vascular endothelial cells [6]. In endothelial cell culture model, ginsenoside-Rg1 is capable of regulating VEGF through activation of Glucocorticoid Receptor (GR) and vascular modeling via endothelial nitric oxide synthease (eNOS) [5-10]. Whether ginsenoside-Rg1 could regulate vascular endothelial signaling under hyperoxic condition needs to be ascertained.

Literatures concerning the application of ginsenoside-Rg1 in protection of hyperoxia-induced vascular endothelium injury are limited. We hypothesize that the treatment of ginsengoid-Rg1 could involve in vascular endothelium signaling under hyperoxic condition. The vascular endothelial oxidative stress and cellular apoptosis were carefully evaluated.

Methods

Study design

In order to evaluate the effects of ginsenoside-Rg1 on HUVECs under hyperoxia, various concentrations of ginsenoside-Rg1 were added to endothelial cells cultured under normoxia and hyperoxia in the experiment period. Cell proliferation and protein production will be measured. In clinical condition, Fick's first law is used to describe the exchange of oxygen in the lung capillaries, where oxygen passes from the alveoli through the alveolar membrane and entering the blood stream. Fick's law can also be applied to cell culture in a similar manner if the oxygen concentration at the air-culture medium interface is considered similar to the oxygen concentration of gas at the surface of the alveolar membrane, and oxygen right above the cell monolayer is paralleled to the oxygen concentration in the blood stream [11]. Thus the culture atmosphere was set at 60% O₂, which is similar to the oxygen percentage administrated to preterm infants with respiratory distress syndrome.

Cells

Primary human umbilical vein endothelial cells (HUVEC, CRL-1730TM) were used in our study. The cells were purchased from ATCC (Manassas, VA). Culture medium M199 and Endothelial Cell Growth Supplement (ECGS) were purchased from Sigma. Fetal Bovine Serum (FBS) was from Gibco.

Ginsenoside-Rg1

Experimental reagent ginsenoside-Rg1 is a reference compound, with purity around 97.7%, purchased from the Division of Chinese Material Medica and Natural Products, National Institute for the Control of Pharmaceutical and Biological Products, Ministry of Public Health, China. A stock solution of Ginsenoside-Rg1 50 mM was prepared in sterile double distilled H₂O and stored at -80°C.

Antibodies

β-actin was used as a loading control. Anti-β-actin antibody was obtained from Millipore (Temecula, CA). Antibodies for VEGF, GR, GPx (Glutathione Peroxidase), Bax, Cyt c (cytochrome c), PARP (Poly ADP-ribose polymerase), caspase 3 were purchased from Cell Signaling Technology, Beverly, MA, USA. Cell proliferation was measured by MTT colorimetric assay (CytoSelectTM MTT Cell Proliferation Assay, CELL BIOLABS, INC.).

Cell Culture

HUVECs were cultured in medium M199 with 10% FBS, 25µg/ml ECGS, and 1% penicillin/streptomycin in an incubator at 37°C. After seeding, the cells were stabilized in incubator under room air over night before experiment. In the experiments, cells were divided into normoxia and hyperoxia groups. In normoxia group, cells remained in incubator supplied with 95% room air (approximately 20% O₂) and 5% CO₂. In hyperoxia group, cells were transferred to a chamber supplied with mixed air containing 60% O₂, 35% N₂, and 5% CO₂. Culture medium was saturated with mixed air previously before experiments. The duration for experiments were 24, 48, and 72 hours after cells were exposed to normoxia or hyperoxia. In hyperoxia group, NexBiOxy Hypoxia/Hyperoxia System (NexBiOxy, Taiwan) was used to monitor and maintain the oxygen concentration in the culture chamber.

Ginsenoside-Rg1 administration

According to references, 0, 75, 150, 300nM of ginsenoside-Rg1 were used in our experiments [6]. Various concentrations of ginsenoside-Rg1 were prepared and added to the medium just before experiment. Medium was changed every day to keep ginsenoside-Rg1 within half-life period.

Western blot

HUVECs were washed in phosphate-buffered saline and then extracted with lysis buffer (50 mM Tris-HCl, 150 mM NaCl, 1% Triton, 0.1% SDS, and 0.5% sodium deoxycholate) and protease inhibitors. Lysates were centrifuged at 12,000 rpm for 5 min, and the resulting supernatant was collected. The extracted protein was quantized by protein assay. Aliquots (25 µg) of cellular lysate was separated by 8% sodium dodecyl sulphate-polyacrylamide gel electrophoresis and transferred to polyvinylidene fluoride (PVDF) membrane. After blocking with 5% BSA, blots were incubated with primary antibodies and then corresponding secondary antibodies. An enhanced chemiluminescence kit (Amersham, Piscataway, NJ) was used for immunodetection. The protein bands were quantified using the Image J software (NIH, Bethesda, MD).

Statistical analysis

Each experiment was repeated for at least three times. Data were expressed as means \pm S.D. Statistical comparisons were carried out

by one-way ANOVA for multiple groups or t test for two groups. Significance was accepted at the p \leq 0.05 level and was marked with * in the figures.

Results

HUVECs proliferation under room air and 60% O,

We firstly assessed the impact of oxygen concentration on HUVEC proliferation in response to ginsenoside-Rg1. Under normoxic conditions (room air), HUVECs exhibited a time-dependent increase in cell number following Rg1 stimulation (Figure 1A), consistent with its reported pro-proliferative properties. In contrast, exposure to hyperoxia (60% O2) resulted in a significant reduction in HUVEC proliferation upon Rg1 treatment (Figure 1B). These findings suggest that ginsenoside-Rg1 may exacerbate oxygen-induced cytostatic effects, impairing endothelial cell expansion under hyperoxic stress, while exerting no inhibitory influence under physiological oxygen levels.

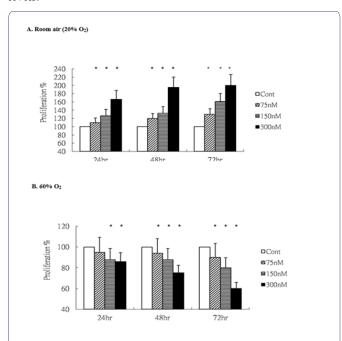


Figure 1: Rg1 downregulates HUVECs proliferation under 60% O2. HUVECs exposed to room air or 60% O2 in paired chambers and treated with various concentrations of Rg1 for 24, 48 and 72hr, respectively. Cell proliferation was determined by colorimetry based on the uptake of MTT. (A). Under room air condition, the total number of HUVECs increased gradually in a time dependent fashion in response to Rg1 stimulation. (B). Under 60% O2 condition, the total number of HUVECs was significantly decreased in response to increasing concentrations of Rg1 (75, 150, and 300nM) stimulation. *P<0.05 vs. Control. n=5.

Levels of VEGF

Next, we evaluated VEGF expression to elucidate its regulatory role in vascular endothelial cell proliferation under both normoxic and hyperoxic conditions. Under room air, the levels of VEGF were significantly elevated in response to ginsenoside-Rg1 treatment in a time-dependent fashion (Figure 2A). Under 60% $\rm O_2$ condition, ginsenoside-Rg1 significantly decreased the levels of VEGF in a time-dependent fashion compared with control (Figure 2B). The inhibition of VEGF expressions indicating the ginsenoside-Rg1 could damper the potential vasogenesis under hyperoxic condition.

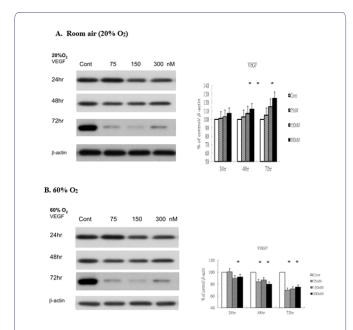


Figure 2: Rg1 does not induce VEGF production in HUVECs under 60% O2. HUVECs exposed to room air or 60% O2 in paired chambers and treated with various concentrations of Rg1 for 24, 48 and 72hr, respectively. The levels of VEGF were determined by Western blot. b-actin was used as the loading control. The signal intensities were determined by densitometry. Data are shown as mean \pm SD of three independently experiments. (A). Under room air condition, the levels of VEGF were significantly elevated in response to Rg1 stimulation. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) increased the level of VEGF. The stimulatory effect of Rg1 on VEGF production on HUVECs was time-dependent. (B). Under 60% O2 condition, Rg1 significantly decreased the levels of VEGF compared with control. The inhibitory effect of Rg1 on decreased VEGF production was time-dependent. *P<0.05 vs. Control.

Levels of GR

To explore the potential involvement of glucocorticoid signaling in Rg1-mediated endothelial responses, we next examined the expression of the Glucocorticoid Receptor (GR) under both normoxic and hyperoxic conditions. Under room air, GR levels were significantly elevated following ginsenoside-Rg1 treatment (Figure 3A). Similarly, under 60% O₂, Rg1 induced a marked increase in GR expression at 72 hours compared with control (Figure 3B). This upregulation suggests that ginsenoside-Rg1 may enhance vascular signal transduction pathways mediated by GR, particularly under hyperoxic stress.

Levels of GPx

To investigate the potential antioxidant response elicited by ginsenoside-Rg1, we analyzed cytosolic glutathione peroxidase (GPx) levels in HUVECs following treatment. Under normoxic conditions, GPx expression remained unchanged in response to Rg1 (Figure 4A). In contrast, under hyperoxic exposure (60% O₂), GPx levels were significantly upregulated following Rg1 treatment (Figure 4B). Notably, increasing concentrations of Rg1 (75, 150, and 300 nM) induced a dose- and time-dependent elevation in GPx expression. These findings suggest that ginsenoside-Rg1 may enhance antioxidant defense mechanisms under hyperoxic stress, potentially contributing to redox regulation in vascular endothelial cells.

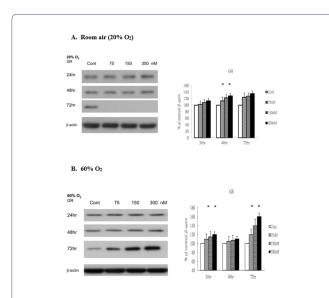


Figure 3: Rg1 induces GR production in HUVECs under 60% O2. HUVECs exposed to room air or 60% O2 in paired chambers and treated with various concentrations of Rg1 for 24, 48 and 72hr, respectively. The levels of GR (glucocoticoid receptor) were determined by Western blot. β-actin was used as the loading control. The signal intensities were determined by densitometry. Data are shown as mean \pm SD of three independently experiments. (A). Under room air condition, the levels of GR were significantly elevated in response to Rg1 stimulation. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) increased the levels of GR. (B). Under 60% O2 condition, Rg1 elevated the levels of GR compared with control. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) elevated significantly the level of GR at 72hr. *P<0.05 vs. Control.

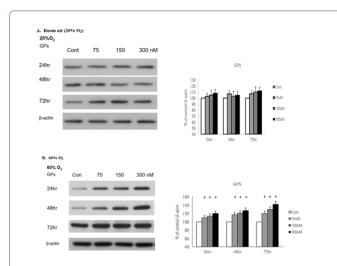


Figure 4: Rg1 induces glutathione peroxidase production in HUVECs under 60% O2. HUVECs exposed to room air or 60% O2 in paired chambers and treated with various concentrations of Rg1 for 24, 48 and 72hr, respectively. The levels of glutathione peroxidase (GPx) were determined by Western blot. β-actin was used as the loading control. The signal intensities were determined by densitometry. Data are shown as mean ± SD of three independently experiments. (A). Under room air condition, the levels of GPx did not change in response to Rg1 stimulation. (B). Under 60% O2 condition, the levels of GPx were significantly elevated in response to Rg1 stimulation. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) increased the level of GPx. The stimulatory effect of Rg1 on GPx production was time-dependent. *P<0.05 vs. Control.

Levels of Bax and Cytochrome c

Because GPx levels remained unchanged under normoxic conditions, implying minimal oxidative stress encounted, we subsequently examined the oxidative stress markers Bax and cyt c under hyperoxic conditions to evaluate whether ginsenoside-Rg1 exacerbates oxidative stress signaling. Under 60% $\rm O_2$ condition, the levels of Bax (Figure 5A) and Cyt c (Figure 5B) increased significantlyin a dose- and time-dependent manner when compared with those in control.

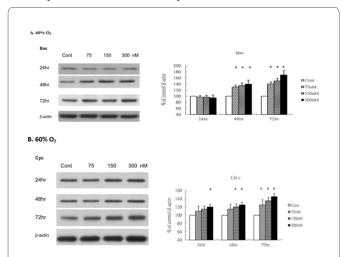


Figure 5: Rg1 increases Bax and cytochrome c (Cyt c) production in HUVECs under 60% O2. HUVECs exposed to room air or 60% O2 in paired chambers and treated with various concentrations of Rg1 for 24, 48 and 72hr, respectively. The levels of Bax and cytochrome c (Cyt c) were determined by Western blot with β-actin used as the loading control. The signal intensities were determined by densitometry. Data are shown as mean \pm SD of three independently experiments. (A). Under 60% O2 condition, the levels of Bax were significantly elevated in response to Rg1 stimulation. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) increased the level of Bax. The stimulatory effect of Rg1 on Bax production was time-dependent. (B). Under 60% O2 condition, the levels of Cyt c were significantly elevated in response to Rg1 stimulation. Treatment with 300nM Rg1 increased the level of Cyt c. The stimulatory effect of Rg1 on Cyt c production was time-dependent. *P<0.05 vs. Control.

Levels of active Caspase 3 and cleaved Poly ADP-Ribose Polymerase (PARP)

We next sought to determine whether vascular endothelium apoptosis could be influenced after ginsenoside-Rg1 treatment, the Western blot was performed. Under 60% $\rm O_2$ condition, the levels of active Caspase 3 (Figure 6A) and cleaved PARP (Figure 6B) increased significantly when compared with those in control. The levels of active caspase 3 (cleaved 17 and 19 kDa) were significantly increased under 60% $\rm O_2$. Similar to active caspase 3, immunoblotting analysis of cleaved PARP showed that the C-terminal 85 kDa PARP apoptotic fragment was significantly generated under 60% $\rm O_2$, indicating vascular endothelium apoptosis has upregulated after ginsenoside-Rg1 treatment.

Discussion

Ginseng has been used as a tonic remedy in traditional complementary medicine for over 2000 years. In recent years, many beneficial effects of ginsenoside-Rg1, one of the major active ingredients of ginseng, have been reported. The effects include increasing glucose

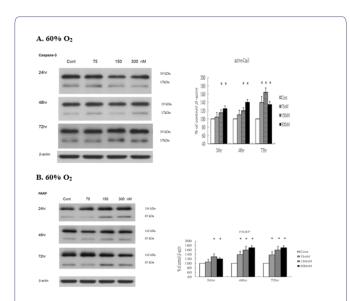


Figure 6: Rg1 increases proapoptotic active caspase 3 and Poly (ADP-ribose) polymerase (PARP) production in HUVECs under 60% O2. HU-VECs exposed to room air or 60% O2 in paired chambers and treated with various concentrations of Rg1 for 24, 48 and 72hr, respectively. The levels of active caspase 3 and PARP were determined by Western blot with β -actin used as the loading control. The signal intensities were determined by densitometry. Data are shown as mean \pm SD of three independently experiments. (A). Under 60% O2 condition, the levels of caspase 3 were significantly elevated in response to Rg1 stimulation. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) increased the level of active caspase 3. The stimulatory effect of Rg1 production was time-dependent. (B). Under 60% O2 condition, the levels of PARP were significantly elevated in response to Rg1 stimulation. Treatment with increasing concentrations of Rg1 (75, 150, and 300nM) increased the level of PARP. The stimulatory effect of Rg1 on PARP production was time-dependent. *P<0.05 vs. Control.

uptake, relieving oxidative stress, suppressing adipocyte development and possible neuroprotective role [12]. It is a potent proangiogenic factor of vascular endothelial cells and stimulator of VEGF expression in human umbilical vein endothelial cells [6]. We hypothesize that treatment with ginsenoside Rg1 may participate in modulating vascular endothelial signaling under hyperoxia.

In this study, Human Umbilical Vein Endothelial Cells (HUVECs) were cultured under normoxic (room air) and hyperoxic (60% O₂) conditions, followed by administration of ginsenoside Rg1. Endothelial proliferation, oxidative stress, and apoptosis were systematically assessed. Under normoxia, Rg1 significantly enhanced HUVEC proliferation and VEGF expression in a time- and concentration-dependent manner, whereas these effects were markedly attenuated under hyperoxic exposure. Ginsenoside Rg1 has been reported to activate the Glucocorticoid Receptor (GR), a pivotal regulator involved in initiating angiogenic signaling pathways [5,13]. Consistent with these findings, our data demonstrate that Rg1 treatment upregulated GR expression in HUVECs under both normoxic and hyperoxic conditions, with the highest GR levels observed following 72-hour exposure to hyperoxia. Previous studies have shown that the proangiogenic effects of ginsenoside Rg1 are mediated through VEGF production via glucocorticoid receptor (GR)-dependent signaling pathways in HUVECs [5,7,13]. In contrast, our findings under hyperoxic conditions revealed increased GR expression accompanied by reduced VEGF levels following Rg1 treatment, suggesting activation of GR-dominant signaling independent of VEGF-mediated angiogenesis. Notably, Rg1 has been identified as a GR agonist capable of inducing nitric oxide production via endothelial nitric oxide synthase [9], and exerting anti-inflammatory effects through GR-mediated pathways [14,15]. These observations imply that under hyperoxia, GR-dominant signaling triggered by Rg1 may contribute primarily to vascular anti-inflammatory responses rather than angiogenic activity [16,17].

Glutathione peroxidase (GPx), a vital antioxidant enzyme within the selenoprotein family, mitigates oxidative damage by catalyzing the reduction of lipid hydroperoxides to alcohols and converting hydrogen peroxide into water. In our study, HUVECs exposed to hyperoxia and treated with ginsenoside Rg1 exhibited a significant, timeand dose-dependent increase in GPx expression. This upregulation suggests that Rg1 may activate intrinsic antioxidant defense mechanisms to counteract hyperoxia-induced oxidative stress. Supporting this interpretation, previous studies have also demonstrated that Rg1 confers cytoprotection against oxidative injury through modulation of the glutathione system [18-20].

Hyperoxia has been shown to induce cell death in vascular endothelial cells, primarily via apoptotic pathways driven by the accumulation of reactive oxygen species and inflammatory mediators [21-25]. In our study, ginsenoside Rg1 treatment under hyperoxic conditions resulted in a concentration-dependent increase in the expression of Bax, active caspase-3, cytochrome c (Cyt c), and PARP, indicating activation of intrinsic apoptotic signaling.

Bax, a pro-apoptotic member of the Bcl-2 protein family localized to the outer mitochondrial membrane, plays a pivotal role in initiating apoptosis through the release of cytochrome c (Cyt c) and subsequent caspase activation [26,27]. Our findings suggest that higher concentrations of ginsenoside Rg1 under hyperoxic conditions may exacerbate apoptosis in HUVECs, as evidenced by elevated levels of Bax and Cyt c. To further assess whether Cyt c release reflects hyperoxia-induced apoptotic progression, we examined the cleavage of poly (ADP-ribose) Polymerase (PARP), a hallmark of programmed cell death. Notably, increased levels of the cleaved 85 kDa PARP fragment were observed, consistent with activation of a PARP-dependent apoptotic pathway involving apoptosis-inducing factor translocation [28]. The dose-dependent activation of PARP in response to Rg1 under hyperoxia underscores its potential to aggravate vascular endothelial cell death.

Under hyperoxic conditions (60% O_2), ginsenoside Rg1 did not promote vascular endothelial cell proliferation or VEGF expression, indicating its limited capacity to counteract hyperoxia-induced endothelial injury. Additionally, the dose-dependent upregulation of pro-apoptotic markers including Bax, cytochrome c, active caspase-3, and cleaved PARP suggests that Rg1 may exacerbate apoptotic signaling under oxidative stress. Conversely, the observed elevation of GR and GPx points to a potential involvement of Rg1 in modulating vascular redox signaling and antioxidant defense mechanisms.

Conclusion

These findings highlight the dualistic nature of ginsenoside Rg1's bioactivity in hyperoxic environments and underscore the need for further mechanistic studies to clarify its role in endothelial survival and stress adaptation.

Ethics Approval and Consent to Participate

Not applicable. Since there is no patient involvement, no ethics approval and consent required.

Funding Declaration

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Consent for Publication

Not applicable.

Availability of Data and Materials

All datasets generated and analyzed during the current study are not publicly available because not yet published, but are available from the corresponding author on reasonable request.

Competing Interests

There are no financial or non-financial competing interests.

Authorship Contribution Statement

Ren-Huei Fu: research idea, literature review, protocol design, experiment processing, data analysis and discussion, manuscript writing. Chi-Nan Tseng: literature review, protocol design, data analysis and manuscript discussion. Cih-Yi Yen & Yu-Hsueh Cho: protocol design, experiment processing, data collecting. Yen Chu: research idea, literature review, protocol design and improvement, experiment guiding and processing, data analysis and discussion, manuscript writing and refinement.

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